

NYSDOH Environmental Laboratory Approval Program – PLM Checklist

LAB ID:	LABORATORY NAME:
DATE:	ASSESSOR NAME:

**BULK ASBESTOS AND SURFACING MATERIAL CONTAINING VERMICULITE SAMPLES BY POLARIZED-LIGHT MICROSCOPY**

**Method Number:**

**SOP Number:**

**Revision Number:**

**SOP Date:**

**Personnel records observed** (including seasonal if applicable):

**Data records observed:**

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	Y	N	NA	Comment	Code
<b>BULK ASBESTOS AND SURFACING MATERIAL CONTAINING VERMICULITE SAMPLES BY POLARIZED-LIGHT MICROSCOPY</b>					
<b>I. Analytical Method (G1004)</b>					
Item 198.1 or EPA 600/M4/82/020 is for Friable Bulk Samples, and Item 198.6 is for Non-Friable Organically Bound Bulk Samples. NYS does not allow visual estimation (EPA 600/R93/116). Item 198.8 is for Surface material containing Vermiculite Bulk Samples.					
A. Are necessary charts and tables available to analyst (e.g., McCrone 1989 or Su 1994 or 2009 dispersion staining table)?					G1012
B. Does the lab maintain a list of non-asbestos fibers that can be confused with asbestos? (Note: This could be a poster, SOP, etc.)					G1245
a. Does the list include optical properties that disqualify each fiber being identified as asbestos?					G1246
C. Does the lab have a textbook or reference book on mineralogy or crystallography (e.g., McCrone 1980; McCrone 1988; Deer, Howie, and Zussman 1996, Shelly 1975)?					G1247
<b>II. Polarized-Light Microscope (G1016)</b> (Section 3 of Items 198.1 and 198.6 and Section 1.5 and 1.6 of EPA 600/M4/82/020)					
A. Is the PLM equipped with the following:					
a. substage polarizer?					G1020
b. analyzer oriented perpendicular to substage polarizer?					G1024
c. eyepiece with a fixed crosshair aligned in direction of polarizer?					G1028
d. 550 nm (first-order red) retardation/compensator plate at 45° to the polarizer?					G1032
e. graduated rotating stage (360° in 1° increments)?					G1036
f. focusable condenser with centerable iris diaphragm?					G1040
g. low (3.2-10X) and high, dry (30-50X) magnification objective? (Sec. 3.19.7 of Item 198.1) (Sec. 1.5.1 of EPA 600/M4/828/020 states "Objective lenses: 10X, 20X, and 40X or near equivalent.)					G1044
h. eyepiece of ≥ 8X magnification?					G1048
i. Chalkley point-count reticle? (optional)					G1049
<b>III. Equipment and Supplies (G1052)</b> (Section 3 of Items 198.1, 198.6, and 198.8 and Section 1.5 and 1.6 of EPA 600/M4/82/020)					
A. Does the lab have the following equipment/material:					
a. laminar-flow hood or negative pressure glove box with HEPA filtration?					G1056
b. low-power (10-45X) stereobinocular microscope with external source for gross examination?					G1060
c. forceps, dissecting needles, probes, scalpel or razor blades for manipulating bulk samples?					G1064
d. smooth removable substrates (glassine paper or clean glass plate) as surfaces for manipulating bulk samples?					G1068
e. <b>homogenization equipment</b> that includes:					
1. mortar and pestle?					G1080
2. mini-blender (approximately 30-mL capacity), liquid-nitrogen mill, or Wiley mill?					G1084
g. <b>filtration apparatus</b> for polycarbonate filters?					G1432
1. 0.4-µm-pore polycarbonate filters?					G1428
2. petri dishes (50 mm diameter) and lids?					G1429
h. <b>muffle furnace</b> capable of sustained operation at 500°C?					G1096

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1. crucibles (bottom and lid)?					G1436
2. one of the following instruments or materials capable of calibrating a muffle furnace at 480°C: a.) high-temperature thermometer with range to at least 500°C and with subdivisions of 5°C or less, b.) melting-point solids with capability of differentiating 5°C differences between 400°C and 500°C, or c.) potentiometer capable of differentiating 5°C differences between 400°C and 500°C?					G1095
i. dessicator?					G1443
j. analytical balance with sensitivity of 0.0001g?					G1445
k. concentrated hydrochloric acid (reagent grade)?					G1444
l. reagent-grade dilute acetic or hydrochloric acid? (For <b>Bulk</b> , EPA 600/M4/82/020, Section 1.5.1)					G1104
m. surfactant such as sodium metaphosphate or aerosol OT?					G1108
n. heat lamp, slide warmer, or drying oven?					G1109
o. ultrasonic bath?					G1448
p. filtered (0.1-µm) distilled water or deionized water?					G1452
q. calibrated thermometer with range of 0 to 50°C and readability of ±1°C?					G1098
r. microscope slides (75 mm X 25 mm)?					G1072
s. (whole) cover glasses (22 mm X 22 mm)?					G1076
t. marker for labeling slides?					G1077
<b>B. Does the lab have the following reference materials:</b>					
a. NIST SRM 1866a (Common Commercial Asbestos – chrysotile, amosite (grunerite), crocidolite (riebeckite), and synthetic glass fiber)?					G1228
b. NIST SRM 1867 (Uncommon Commercial Asbestos – anthophyllite, tremolite, and actinolite)?					G1244
c. at least 10 different Vinyl Asbestos Tiles (VAT) references that have been analyzed by an ELAP-certified TEM lab?					G1456
1. at least 2 of the verified standards are negative for asbestos?					G1464
2. at least 2 of the verified standards have asbestos concentrations between 1 and 10%?					G1468
3. at least 2 of the verified standards have asbestos concentrations greater than 10%?					G1472
d. permanent mount of NIST amosite in refractive index oil with $n_d = 1.680$ ?					G1223
e. a complete set of RI oils ranging from $n_d = 1.49$ to $1.72$ in intervals $\leq 0.005$ ?					G1224
f. either a solid RI calibration material (e.g., Cargille glass) or a refractometer capable of an accuracy of $\pm 0.004$ .					G1252
<b>Additional equipment and supplies for Surfacing Material Containing Vermiculite Bulk Sample Analysis: (G1600)</b> (Section 3 of Item 198.8)					
<b>C. Does the lab have the following equipment/material and reference materials:</b>					
a. centrifuge, capable of 3600 rpm and accommodating at least four 15 mL centrifuge tubes?					G1088
b. glass or polypropylene, centrifuge tubes, 15 mL capacity?					G1601
c. at least 4 different SM-V specimens analyzed by outside lab?					G1602
d. at least 2 negative (non-ACM) standards?					G1603
e. at least 2 positive (ACM) standards?					G1604

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	Y	N	NA	Comment	Code
f. magnetic stirrer (Teflon coated, 5 cm in length) and magnet?					G1605
g. Sink-Float® Standard (2.75±0.005 g/cc at 23°C)?					G1606
h. water aspirator?					G1607
i. Erlenmeyer flasks (conical, 250 mL)?					G1608
j. 25 mm and 47 mm diameter glass vacuum filtration assembly ?					G1609
k. polycarbonate filters (0.4-0.8 um, 47 mm diameter)? Note: 25 mm is allowed when there is a small amount of centrifugate.					G1610
l. porcelain or glass Buchner funnel (240 mL)?					G1611
m. Whatman 40 cellulose filters (90 mm diameter)?					G1612
n. 25 mm and 47 mm diameter mixed esters of cellulose filters (0.22 um porosity)?					G1613
o. heavy liquid (either an aqueous solution of lithium metatungstate or sodium polytungstate)?					G1614
p. reagent grade ethanol or methanol?					G1615
q. cotton applicator swabs (to remove material from upper part of centrifuge tubes)?					G1616
<b>IV. Sample Preparation – NOB Bulk Samples and Friable Sample Problem Matrices (G1112)</b>					
A. Are samples homogenized when necessary?					G1116
a. Are at least 4 subsamples prepared and mounted? (Item 198.1)					G1118
b. Are at least 8 subsamples prepared and mounted? (EPA 600/M4/82/020, Section 1.7.2.4)					G1617
<b>Refer to the “Matrix Modification” Section (XI) of this checklist.</b>					
B. Are samples acid treated when necessary?					G1120
C. Are samples dispersed with surfactant when necessary?					G1124
D. Are samples ashed when necessary at 480°C until mass stabilizes (1-12 hours)?					G1128
E. Are layers in layered samples analyzed individually?					G1130
<b>V. Sample Amount, Storage, and Preparation – Surfacing Material Containing Vermiculite Bulk Samples (G1700)</b>					
Gravimetric Reduction (Section 4.2.1)					
A. Is a minimum weight of 3 grams used for analysis?					G1701
B. Did laboratory notify its clients that a minimum of 10 grams of sample is required?					G1702
C. Does the laboratory keep any unused portion of sample for a period of no less than 90 days from the date the report is transmitted to the client?					G1703
D. Are samples ashed at 485 ±5°C for at least 10 hours?					G1704
Acid Treatment (Section 4.2.2 and 4.2.3)					
E. Is the sub-sample acid treated according to Section 4.2.2. and 4.2.3?					G1705
F. Is the floatable material in the petri dish dried until a stable weight is achieved (< 3% difference in weight)?					G1706
(Using Erlenmeyer flask, stirring rod, 2 M HCl, 0.1 um filtered water, and magnet. Sample is stirred for 15 minutes. Removing any floatable material. Repeating rinses at least 3 times.)					
Filtration (Section 4.3)					
G. Is the remaining materials (liquids and solids) collected during acid treatment filtered according to Section 4.3?					G1707

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	Y	N	NA	Comment	Code
(Using glass filtration apparatus, 0.4-0.8 um polycarbonate filter, and 0.1 um filtered water. Repeating rinse at least 2 times.)					
H. Is the filtered material in the petri dish dried until a stable weight is achieved (< 3% difference in weight)?					G1708
<b>VI. PLM examination for Chrysotile (Section 6), Determination of Amphibole Asbestos Using Heavy Liquid Centrifugation (7), PLM examination for Amphibole Asbestos (8), and Calculations (9) – Surfacing Material Containing Vermiculite Bulk Samples</b>					
Section 6 (G1800)					
For the examination of chrysotile, ...					
A. Are eight slides prepared from the residue using high dispersion liquid of RI 1.630 or 1.680?					G1801
B. Is each slide scanned using crossed polars with a 550 nm compensator plate to determine if structures morphologically consistent with chrysotile are present?					G1802
C. If no structures are detected, are zero chrysotile points and 50 occupied points assigned to each slide?					G1803
D. If structures are detected, is at least 1 additional slide using RI 1.550 oil prepared and are at least 4 structures positively identified as per Section 5?					G1804
For the quantitation of chrysotile, ...					
E. Does the lab use the 400 point count method with 50 non-empty points per slide?					G1805
F. Does the lab use the original 8 slides (in RI 1.630 or 1.680 oil)?					G1806
(Chalkley point-count reticle is not allowed.)					
Section 7 (G1825)					
For the determination of amphibole asbestos concentration using heavy liquid centrifugation, ...					
A. Is the residue properly dried and weighed? (i.e., If the residue has been exposed to room air for > 1 hour, the residue must be placed back in the oven or desiccator for a minimum of 1 hour before weighing.)					G1826
B. Is the residue properly divided and transferred between 2 centrifuge tubes?					G1827
C. Is the heavy liquid calibrated as per Appendix D?					G1828
E. Is the residue properly dispersed throughout the heavy liquid? (i.e. using a glass rod, adding at least 10 mL of heavy liquid to bring the liquid level up to 2 cm from the top of each tube)					G1829
F. Are the tubes properly centrifuged? (i.e. Centrifugation times vary depending on the dimensions and rotation speed of the particular centrifuge. Refer to Appendix E.)					G1830
G. Is the heavy fraction properly washed? (i.e. washing 5 times with 2 mL reagent water)					G1831
H. Is the centrifugate properly washed to remove the heavy liquid? (i.e. using 5 mL 0.1 um water; one wash)					G1832
Section 8 (G1850)					
For the examination and quantitation of amphibole, ...					
A. Are eight slides prepared from the residue using high dispersion liquid of RI 1.630?					G1851
B. Is the whole area of each slide scanned at a 100x magnification as per Section 5?					G1852
C. If no amphibole asbestos is observed, are 50 occupied points and zero asbestos points assigned to each slide?					G1853
D. If amphibole asbestos is observed, is scanning discontinued and 400 point count started?					G1854
E. Does the lab calculate the concentration of amphibole asbestos in the centrifugate and in the original sample as per Appendix B and Section 9?					G1855

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	Y	N	NA	Comment	Code
Section 9 (G1875)					
For the calculation of chrysotile, does the lab follow the calculations in Appendix B and Section 9?					G1876
For the calculation of amphibole, does the lab follow the calculations in Appendix B and section 9?					G1877
For the calculation of total asbestos content, does the lab follow the calculations in Appendix B and Section 9?					G1878
<b>VII. Analytical Records (G1132)</b>					
A. Does the analysis sheet include/indicate the following:					
a. analyst's signature or initials and date of analysis?					G1136
b. sample ID number?					G1140
c. gross description of material including color, homogeneity and texture?					G1144
d. disqualifying optical property for each non-asbestos fiber identified?					G1177
e. matrix reduction? (See Section X, too.)					G1148
f. at least four subsamples are prepared and mounted? (Item 198.1)					G1180
g. at least eight subsamples are prepared and mounted? (EPA 600/M4/82/020)					G1152
h. use of EPA 600/M4/82/020 and/or ELAP Item 198.1 point count methods?					G1184
i. tally of points for each type of asbestos?					G1188
j. original quantitation results that are based on point counting?					G1192
B. Does the analysis sheet for <b>SM-V</b> capture the following (in addition to items a.-j. noted in Section VII, A, above):					
a. confirmation of vermiculite by stereo binocular microscope?					G1193
b. matrix reduction (mass of original sub-sample, mass after ashing, mass after acidification, mass after acid float, mass of centrifugate)?					G1148
C. For each type of asbestos type identified, is the following recorded:					
a. morphology?					G1156
b. birefringence?					G1160
c. angle of extinction?					G1164
d. sign of elongation?					G1168
e. RI (to the nearest 0.004) for fiber length (parallel)?					G1172
f. RI (to the nearest 0.004) for fiber width (perpendicular)? (Note: Fiber width and fiber length should be different.)					G1176
<b>VIII. Calibration Records (G1248)</b>					
A. Are there records of the following:					
a. <b>quarterly or next use, whichever is less frequent</b> , calibration of refractive index oils to within 0.004?					G1256
b. calibration of RI oils to within 0.004 when a new container is opened?					G1257
c. <b>daily or next use</b> alignment of PLM?					G1260
d. <b>monthly</b> determinations of dispersion-staining or Becke-line colors from the lab's permanent amosite mount?					G1264
e. <b>semi-annual</b> measurements of HEPA-ventilated enclosure(s) demonstrating a face velocity of at least 75 fpm?					G1268
f. <b>quarterly</b> calibrations of muffle furnace in the range 450-480°C?					G1270

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	Y	N	NA	Comment	Code
g. room temperature being checked <b>daily or next use</b> ?					G1271
<b>IX. Quality Control and Personnel Records (G1376)</b>					
A. Have all QC analyses been performed and evaluated before final reports are submitted to clients?					G1301
B. Are QC samples submitted blindly to the original analyst so that the analyst is unaware that the sample will be reanalyzed? (Note: A random number generator could be used for 1-person labs.)					G1303
C. For intra-analyst precision, does the original analyst reanalyze <b>at least 2%</b> of blind QC samples? (NELAC 2003 Ch. 5 App. D.6.2.13.a.)					G1305
a. For single analyst labs, is <b>at least 1 out of every 10</b> blind QC samples reanalyzed? (NELAC 2003 Ch. 5 App. D.6.2.13.a.)					G1304
D. For inter-analyst precision, does a different analyst reanalyze <b>at least 6.7%</b> of blind QC samples given to the original analyst? (NELAC 2003 Ch. 5 App. D.6.2.13.b.)					G1307
E. Do QC reanalyses include complete and independent reparation and analysis of the sample?					G1309
F. Are R-bar charts showing intra- and inter-analyst precision kept up-to-date for each analyst?					G1310
a. Are records of each analyst's replicate and duplicate analyses kept?					G1388
b. Is corrective action taken when R values are > 1 or < -1 for inter-analyst QC? (Note: Acceptable range = 1 to -1)					G1311
c. Is corrective action taken when <b>absolute</b> R values are > 1 for intra-analyst QC?					G1313
G. Is <b>at least 1%</b> (1 out of 100) of samples analyzed a standard or reference sample that has been routinely resubmitted? (NELAC 2003 Ch. 5 App. D.6.3.3a. and b.)					G1312
a. For friable materials, does at least 50% of the QC reference samples submitted contain between 1 and 10% asbestos?					G1315
H. Are X-bar charts showing analyst's accuracy kept up-to-date?					G1314
a. Are the records from reference standard analyses kept for each analyst?					G1392
I. Are re-analysis of inter-laboratory QC samples performed <b>at least quarterly</b> or <b>at a rate of 1 sample per 500 routine samples</b> (whichever is less)? (Note: NYSDOH PT studies can be used. Each study includes 4 samples.)					G1317
J. Have all misclassifications (false positives and false negatives) and misidentifications of asbestos types associated with inter-laboratory reanalyses been resolved?					G1320
K. Is at least one non-ACM blank prepared <b>daily</b> or <b>with every 50</b> samples analyzed, whichever is less? (Item 198.1, Section 8.3.2)					G1325
a. Or, is a blank check made <b>after every 20</b> uses of each piece of homogenization equipment? (Item 198.1, Section 8.3.2)					G1323
L. Is at least one non-ACM non-friable material prepared and analyzed <b>with every 20</b> samples analyzed? (Item 198.6, Section 8.3.2)					G1327
M. Do <b>monthly</b> summaries reveal an error rate of <b>less than 1%</b> on the classification of samples?					G1326
N. Are all analysts able to correctly identify the six regulated asbestos types? (Note: The six types are chrysotile, amosite (grunerite), crocidolite (riebeckite), anthophyllite, actinolite, and tremolite)					G1395
O. Are records kept for each analyst outlining resolutions of any QC deficiencies?					G1396
P. Are <b>at least 10%</b> of <b>SM-V</b> analyses re-analyzed?					G1397
<b>X. Results and Reports (G1196)</b>					
A. Do the final results include the type and percentage of each asbestos type?					G1200

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B. Do the final results include the type and percentage of each non-asbestos fiber type?					G1204
C. Is the percentage of asbestos detected rounded off to <b>two significant digits</b> ?					G1230
D. Does a recent client report on an ACM include the following:					
a. color of the bulk sample?					G1208
b. type and percentage of each asbestos type?					G1212
c. type and percentage of each non-asbestos fiber type?					G1216
E. Are results from layered samples reported as separate layers? (Note: Labs can do composites, but lab needs to record results for original layers.)					G1217
F. Are the samples stored in a secure area for <b>at least 60 days</b> after result reporting?					G1218
G. Are samples determined by the laboratory to be NOB and analyzed by Item 198.4 and/or 198.6 clearly noted as NOB material on reports?					G1219
H. Do final results for <b>SM-V</b> include all of the following: % chrysotile and amphibole detected, total % asbestos, % organic fraction and water, % floats, % residue, and % centrifugate?					G1231
<b>XI. Matrix Modification for NOB Bulk Samples and Friable Sample Problem Matrices (G1424)</b>					
A. Do analysis sheets show calculation of percent matrix loss during muffle furnace ashing?					G1476
B. Do analysis sheets show calculation of percent matrix loss during acid digestion?					G1478
C. Are reduced samples cooled in a dessicator prior to weighing?					G1488
D. Are percentages of matrix loss used in calculating final asbestos percentage?					G1480
E. Are <b>inconclusive</b> ( $\leq 1\%$ ) NOB asbestos results by PLM reported with the ELAP required disclaimer? (See disclaimer in Item 198.6, Section 6.3.2.2.)					G1484
F. Is the gravimetric reduction method used to generate the asbestos result?					G1487
<b>XII. ELAP On-Site Audit Materials and Samples (G1328)</b>					
<b>Proficiency Test and Routine Samples</b>					
A. Were proper dispersion-staining colors or Becke lines visible with the lab's amosite slide?					G1332
B. Was complete extinction observed when the lab's amosite mount was viewed with crossed polars?					G1336
C. Did the analyst correctly describe the morphology of the fibers?					G1340
D. Was the color of the sample recorded on the analysis sheet?					G1344
E. Were subsamples taken at random and without preference to fibers?					G1348
F. a. Were at least 4 subsamples prepared and mounted using whole coverslips? (Item 198.1)					G1352
b. Were at least 8 subsamples prepared and mounted using whole coverslips? (EPA 600/M4/82/020)					G1374
G. Was the analyst able to accurately determine if the fiber's refractive index was lower or higher than the initial mounting medium?					G1356
H. Was the analyst finally able to determine the refractive index of the fiber length and width to within 0.004 of the known refractive index?					G1360
I. Was sign of elongation correct?					G1364
J. Was extinction angle correct?					G1368
K. a. Was the Item 198.1 stratified point counting done correctly?					G1372
b. Was EPA 600/M4/82/020 point counting done correctly?					G1375



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Note: The analyst must use a uniform scan pattern when analysis is performed with a multi-point eye piece. (Item 198.1, Section 5.2.2)					
L. Did analyst accurately identify fibrous components?					G1373

**Additional Observations/Notes:**

**Other Useful Information:**

Non-Friable Organically Bound materials (NOB): vinyl asbestos tile (VAT), resilient floor tiles, mastic, asphalt shingles, paint chips, caulking, glazing, etc.

R-bar (Item 198.1 and 198.6, Sec. 8.2.2) - **Inter-Analyst**

$$R = \frac{(A - B)}{\left(\frac{A + B}{2}\right)}$$

R-bar (Item 198.1 and 198.6, Sec. 8.2.1) – **Intra-Analyst**

$$R = \left| \frac{(A - B)}{\left(\frac{A + B}{2}\right)} \right|$$

Where A = result from the analyst being checked and B = result from other analyst for same sample.

Multiple analysts: Inter-analyst, at least 1 per 15 samples and Intra-analyst, at least 1 per 50 samples

Single analyst: Intra-analyst, at least 1 per 11 samples

X-bar (Item 198.1 and 198.6, Sec. 8.2.3)

Accuracy of each analyst shall be monitored by determining percent recovery.

$$\% \text{ Recovery} = \left( \frac{A}{W} \right) \bullet 100$$

Where A = analytical result read by analyst and W = formulated weight for reference standard slide

Disclaimer (from Item 198.6, Sec. 6.3.2.2)

"Polarized-light microscopy is not consistently reliable in detecting asbestos in floor coverings and similar non-friable organically bound materials. Quantitative transmission electron microscopy is currently the only method that can be used to determine if this material can be considered or treated as non-asbestos containing."

Table 1 (from Item 198.1, Table I and Item 198.6, Table I)

Asbestos Type	Color and Morphology	Refractive Index		Sign of Elongation	Extinction Angle
		⊥			
Chrysotile	White to pale green; v. flexible w/ "kinks"; Wavy w/ "knuckles"	1.493-1.559	1.517-1.567	+	; undulose
Amosite	Tan; mod. flexible, but straight bundles; easily splayed ends	1.657-1.686	1.696-1.729	+	;
Crocidolite	Dark blue; flexible; some "kinks"; splayed ends; strongly pleochroic	1.654-1.701	1.668-1.717	-	
Anthophyllite	White to light tan; stiff; ends splayed to blunt	1.596-1.652	1.615-1.722	+	
Tremolite	White to light tan; stiff; large bundles; may have splayed ends	1.599-1.628	1.625-1.655	+	; v. thin fibers or cleavage fragments (≤ 15°)
Actinolite	White to green; stiff; large bundles; may have splayed ends; often pleochroic	1.600-1.668	1.625-1.688	+	; v. thin fibers or cleavage fragments (≤ 20°)

% Asbestos NOB Calculation

$$\% \text{ Asbestos} = \left( \frac{PAM}{OM} \right) \bullet AP$$

Where PAM is mass of residue after furnace and acid treatment (in mg), OM is mass of original subsample (in mg), and AP is mean percentage of asbestos (versus inorganic residue) in final slide preparations.